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## **REVOLUTIONIZING DRUG DEVELOPMENT: THE IMPACT OF GENE THERAPY AND CRISPR TECHNOLOGY ON MODERN MEDICINE**

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### **ABSTRACT**

Gene therapy and CRISPR technology offer a surgical strike for genetic disturbances: they furnish precise modifications of genetic material to treat and potentially cure genetic disorders, cancers, and infectious diseases. It involves the introduction, variation, or removal of genes with a view to correcting genetic defects; somatic and germline edits fall thereunder. CRISPR-Cas9 represents a new class of molecular tools for gene editing that can ensure targeted, potent, efficient, and essentially reproducible DNA modifications, thereby facilitating the design of genetic therapies. The review explains the basic concepts of gene therapy and other ways of delivering genes, FDA-approved gene therapy applications, and workings and applications of the CRISPR-Cas system; it discusses how CRISPR is an adjunct for drug development, personalized medicine, and clinical applications regarding cancer and pathological conditions of interest in neurology and infectious diseases. Without a doubt, retrofitting these novel potentials holds immense therapeutic value; current obstacles, such as off-target performances, delivery efficiencies, and ethical dilemmas, remain significant roadblocks to preventing the widespread acceptance of these technologies. The future direction looks at the refinement of the CRISPR technology, enhancement of gene delivery systems, and application of Artificial Intelligence for a better precision in genetic medicine. Thus, the review describes the enormous promise of gene therapy and CRISPR technologies, while the implementation of these technologies faces a multitude of scientific, ethical, and regulatory hurdles.

## 1. Introduction

### 1.1 Overview of Gene Therapy

Gene therapy cures or prevents disease by correcting genetic defects and includes the introduction, alteration, or elimination of genes within a patient's cells [1]. Somatic gene therapy is one type for non-reproductive cells and does not influence the future generation and germline gene therapy that alters the reproductive cells and raises numerous ethical issues [2]. The majority of the therapeutic genes use viral vectors that make use of engineered viruses, but non-viral vectors have also been used, such as nanoparticles [3]. Gene therapy is used to treat monogenic diseases (sickle cell anemia and cystic fibrosis), cancer, infectious diseases, and neurological disorders [4].

Technological advances have delivered CRISPR-Cas9, cancer CAR-T cell therapy, and RNA-based therapy such as mRNA vaccines [5]. Gene therapy is monitored by the FDA, and considerations of ethics encompass germline editing and universal access [6]. Gene therapy in the future is all about precision medicine, in vivo gene editing, and integrating it with other therapy for enhanced efficiency [7]. While promising, the science of gene therapy will have to deal with scientific, ethical, and safety concerns in its development [8].

### 1.2 Introduction to CRISPR Technology

CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) is a gene-editing technology allowing scientists to edit the organism's DNA with precision and efficiency [9]. Discovered initially as part of the bacterial innate immune system, CRISPR acts by using guide RNA and an

enzyme, usually paired up with Cas9, to target and cut the particular DNA sequences, so unaltered genetic material can be added, removed, modified [10].

First, the CRISPR-Cas9 system works by designing a guide RNA that complements the desired DNA sequence [11]. Then, the Cas9 enzyme employs that guide RNA to verify and modify the target DNA sequence [12]. The DNA is cut in that particular location. That break is then repaired by some of the cell's own add-on repair mechanisms, either introducing new genetic material or disabling a previously harmful gene [13]. CRISPR is simple, powerful, efficient, and versatile, which is the reason why it is a great research, medical, and biotechnological tool [14].

Applications of CRISPR technology range from genetic disorder treatments, developing disease-resistant crops, and researching gene function to its use in redesigning therapies to cure diseases such as cancers, sickle cell, and muscular dystrophies [15]. It threatens to be the technology that might permanently change humanity, especially since it raises ethical questions for germline editing that may affect future generations [16]. Nevertheless, CRISPR is fast transforming genetic research and paving avenues for novel innovations in science and medicine [17].

### 1.3 Significance in Drug Development

It is an extraordinary innovation ending all shackles in drug discovery as it knows how to create new avenues to speed up research, improve precision, and develop completely novel ways of treating patients [18]. Most significantly, its fame hangs on precision gene editing-enabled scientists to really cut

genes and study disease mechanisms for possible drug targets and advanced therapeutic development [19].

**Targeted Identification and Validation:** Scientists use knock-out or selective modification techniques for genes, using CRISPR, to perceive disease phenotypes for identification and validation of potential drug targets with activity, while also reducing the budget and time incurred in early-stage drug discovery [20].

**Modeling Diseases:** CRISPR provides model systems at the cellular and animal level that more closely mimic genetic disease by inserting specific mutations [21]. This model will be revelatory in drug candidate testing and disease progression understanding [22].

**Personalized Medicine:** CRISPR will create medications for patients according to their genetic signatures [23]. One could use CRISPR in modifying cells obtained from patients to cure genetic diseases and cancers in a fine and precise manner [24].

**Developing Gene Therapy:** CRISPR is developing therapies that include replacing or modifying genes that have defects, as with sickle cell anemia or beta-thalassemia, and even a few forms of inherited blindness [25].

**Drug Screening:** One can perform large library screening on CRISPR-edited cell lines for an identification of compounds that highly interacted with specific genetic pathways or mutations [26].

## 2. Fundamentals of Gene Therapy (Table 1)

2.1 Table 1: Types of Gene Therapy (Germline vs. Somatic)

Aspect	Somatic Gene Therapy	Germline Gene Therapy
<b>Target Cells</b>	Non-reproductive (somatic) cells (e.g., skin, muscle, organs)	Reproductive cells (sperm, eggs) or early embryos
<b>Inheritability</b>	Changes are <b>not inherited</b> by future generations	Changes are <b>heritable</b> and passed to offspring
<b>Applications</b>	Treats genetic disorders (e.g., cystic fibrosis, sickle cell anemia), cancers, and other diseases	Potential to eliminate genetic diseases from a family line or introduce beneficial traits
<b>Ethical Concerns</b>	Widely accepted, as it affects only the individual	Highly controversial due to heritability and potential for "designer babies"
<b>Current Status</b>	Approved for clinical use in certain conditions (e.g., inherited blindness, blood disorders)	Not approved for use in humans; largely experimental
<b>Advantages</b>	<ul style="list-style-type: none"> <li>- Does not affect future generations</li> <li>- Proven success in treating specific diseases</li> </ul>	<ul style="list-style-type: none"> <li>- Could permanently eradicate genetic disorders</li> <li>- Prevents transmission of genetic diseases</li> </ul>
<b>Challenges</b>	<ul style="list-style-type: none"> <li>- Ensuring efficient delivery to target cells</li> <li>- Avoiding immune responses</li> </ul>	<ul style="list-style-type: none"> <li>- High risk of unintended consequences (e.g., off-target effects)</li> <li>- Ethical and regulatory hurdles</li> </ul>

Aspect	Somatic Gene Therapy	Germline Gene Therapy
	- Long-term gene expression	
Examples	- CAR-T cell therapy for cancer - Treatment for inherited retinal diseases	- Experimental research in animal models - Theoretical applications in humans

## 2.2 Gene Delivery Methods (Viral & Non-Viral Vectors)

They are viruses modified to introduce genes into their cells, providing very high levels of gene expression efficiency and long-term expression [27]. These include adenovirus, retrovirus, lentivirus, and AAV (adeno-associated viruses); their algos are due to immune responses, limited carrying capacity, and insertional mutagenesis [28]. Treatments of genetic disorders, cancers, and inherited retinal diseases use these vectors [29].

Non-viral delivery systems are liposome, nanoparticles, electroporation, or gene guns and are considered safer, easier and carrying larger payloads of genetics but less efficient and usually transitory [30]. The applications of non-viral delivery are DNA vaccines, cancer therapy, and gene editing [31].

In short, virus vectors deliver efficiently but unsafe treatment, while non-viral vectors deliver safely but ineffectively [32]. The choice between the two vectors depends on the specific application, target tissue, and what one expects from the treatment [33]. Improvements of both methods are ongoing, therefore raise the possibilities of gene therapy [34].

## 2.3 Current FDA-Approved Gene Therapies

The FDA so far has given approvals to a handful of gene therapy treatments. These gene therapy treatments mostly target

infrequent genetic disorders and some cancers. Noteworthy examples include:

Luxturna (voretigene neparvovec): Approved in 2017, Luxturna is used to treat patients with inherited retinal diseases such as Leber congenital amaurosis that are due to mutations in the RPE65 gene [35]. It uses an adeno-associated virus (AAV) vector to deliver a functional copy of the RPE65 gene to retinal cells [36].

Zolgensma (onasemnogene abeparvovec): Approved in 2019, Zolgensma treats spinal muscular atrophy (SMA), a serious neuromuscular disorder with genetically base mutation in the SMN1 gene [37]. It uses an AAV vector to introduce a functional SMN1 gene, which improves muscle function and survival [38].

Kymriah (tisagenlecleucel): Approved in 2017, Kymriah is a CAR-T cell therapy for some specific types of B-cell acute lymphoblastic leukemia (ALL) and large B-cell lymphoma [39]. The patient T cells are modified and targeted to destroy the malignant cancer cells [40].

Yescarta (axicabtagene ciloleucel): Approved in 2017, Yescarta is another CAR-T cell therapy used to treat large B-cell lymphoma [41]. It works similarly to Kymriah in that it reprograms T cells to attack cancer cells [42].

Tecartus (brexucabtagene autoleucel): Approved in 2020, Tecartus is a CAR-T cell therapy for mantle cell lymphoma and B-

cell precursor ALL [43]. It uses genetically engineered T cells to attack cancer cells [44]. Abecma (idecabtagene vicleucel): Approved in 2021, Abecma is a new CAR-T cell therapy against multiple myeloma [45]. It targets the B-cell maturation antigen (BCMA) on myeloma cells [46].

Skysona (elivaldogene autotemcel): approved in 2021, Skysona is a lentiviral-derived gene therapy for cerebral adrenoleukodystrophy (CALD), a rare degenerative neurodisorder [47]. The vector delivers a functional ABCD1 gene into hematopoietic stem cells [48].

Hemgenix (etranacogene dezaparvovec): Approved in 2022, Hemgenix is used to treat hemophilia B by replacing the functional factor IX gene using an AAV vector [49]. This lessens the frequency of required clotting factor infusions [50].

These are really great advances in the treatment of previously untreatable diseases [51]. They bring all the patients with rare genetic disorders as well as some cancers close on the cure[52]. However, they are usually very expensive and require to be administered in a very specialized center [53].

### 3.CRISPR-Cas System: Mechanism and Applications

#### 3.1 CRISPR-Cas9 Mechanism

The **CRISPR-Cas9** is a gene-editing tool in general derived from bacteria [54]. It employs guide RNA (gRNA) as an indirect

approach to a particular DNA sequence where the **Cas9 enzyme** takes up the target[55]. It was known to create a double-strand break. This break is typically repaired by the cell through either **non-homologous end joining (NHEJ)**, which possibly disrupts the gene, or **homology-directed repair (HDR)**, which allows the insertion of precise edits when it has a repair template[56]. CRISPR-Cas9, thus, can be used to knockout, correct, or insert a gene, and it is a very potent tool for research, medicine, or biotechnology [57]. The challenges, such as off-target effects and delivery to the target sites, are still being worked on [58].

#### 3.2 CRISPR Variants (Cas12, Cas13, Base Editing, Prime Editing)

Cas12 - Cleaves DNA, creating sticky ends, with a collateral destruction of ssDNA. For use in gene editing and DNA detection (e.g., SHERLOCK) [59].

Cas13 - Targets RNA, producing collateral degradation of RNA. For use in RNA editing and virus detection [60].

Base Editing - Alters single bases of DNA (A→G, C→T) without cutting. Best suited for very precise gene corrections [61].

Prime Editing - In tiny insertions, deletions, or replacements, pegRNA + reverse transcriptase are utilized to achieve DNA manipulation, avoiding double-strand breaks. Most flexible tool for editing [62].

3.3 Table 2 Applications in Genetic Disorders

CRISPR Variant	Application	Targeted Disorders
Cas9 & Cas12	Gene knockout & correction	Sickle cell anemia, Cystic fibrosis
Cas13	RNA-based therapies	ALS, Huntington's disease
Base Editing	Precise single-base	Genetic blindness, Metabolic



	corrections (A→G, C→T)	disorders
Prime Editing	Complex DNA modifications (insert, delete, replace)	Duchenne muscular dystrophy, Tay-Sachs disease

#### 4. Gene Therapy and CRISPR in Drug Development

##### 4.1 Target Identification and Validation

Target identification-the term itself suggests that it could potentially be gene, protein, or RNA-based, which becomes disease-centric and can thus be topic-specific concerning drug discovery endeavours (Table 2) [63].

Methods are:

CRISPR Screens - Determine essential genes.

Genomics & AI - This can actually help in predicting the promising targets.

Proteomics & Transcriptomics-Analysis of disease pathways.

Image Validation: Proves the target would be a potential target in terms of identification of its role to disease and in druggability.

Techniques:

CRISPR Knockouts and Base Editing - Gene function testing.

RNA Interference (RNAi) - Gene silencing.

Animal Models and Cell Assays - Effect validation.

##### 4.2 Drug Discovery and Screening

##### Drug Discovery and Screening

Drug discovery looks for new therapeutic agents with the ability to achieve a cure for diseases [64]. The study of mechanisms of action of the diseases flowing from there, the choices of possible therapeutic targets, and the design of possible interacting molecules with these targets are of interest [65].

Although drug discovery has been awakening a great many new doors through combinatorial chemistry, genomics, bioinformatics, and CRISPR technology, there indeed is a positive hold of these towards the updating of the drug [66].

Drug discovery by screening large numbers of compounds against a given biological target is termed high-throughput screening (HTS) [67]. This practice will take a lead in identifying lead compounds with suitable therapeutic effects[68]. Furthermore, in this direction, using AI-based in silico models, virtual screening has been developed for the identification of interacting drug targets, thereby reducing laboratory testing burdens [69].

CRISPR screening provides an impetus for drug discovery by enabling genome-wide approaches for finding key genes involved in the disease pathway [70]. On the other hand, CRISPR could serve as a rationale to develop disease models for better validation of drug targets and for testing the efficacy of candidate therapeutics in an accurate and efficient way, thereby speeding the identification of drug candidates with maximum specificity and minimum toxicity [71].

Another name for lead optimization is the next process after selecting lead drug candidates for increasing efficacy, safety, and stability [72]. Preclinical studies are then performed in the cell line and animal models to elucidate parameters leading toward clinical relevance [73]. CRISPR, with

the implementation of drug discovery and their screening concepts, keeps fine-tuning the broader work and will definitely present targeted and viable therapies [74].

#### **4.3 Personalized and Precision Medicine**

Medicine personalized to an individual's genetic profile, lifestyle, and environment rather than a one-size-fits-all method, treatment will be considered [75]. Precision medicine has a subset of medicine personalized to use genetic insights in creating targeted therapies and Gene therapy and CRISPR technology have revolutionized this area by enabling precise genetic modifications for root treatment of diseases [76].

CRISPR focuses on editing gene at the molecular level with high specificity in correcting the mutations causing diseases [77]. Such treatment is designed to repair the faulty genes for long-term or even permanent cure for such genetic disorders as sickle cell disease and cystic fibrosis [78]. In cancer, CRISPR enhances development into such biomedical areas as personalized immunotherapies, like the CAR-T cell therapy, where immunocytes of the patient are engineered to target cancer in a better way [79].

Gene therapy also paves the way for the development of drugs targeted at gene modifications that are responsible for drug metabolism[80]. By this approach, one can tailor drug dosages according to the genetic profile of an individual, minimizing side effects and getting better treatment outcomes [81]. CRISPR-based gene modifications are used to develop personalized medicines for neurological diseases like Huntington's disease and ALS.

In this case, gene editing will prevent the disease progression [82].

As it progresses with research, CRISPR and gene therapy further promise transforming drug development through patient-specific therapies, eliminating the trial and error in treatment selection, and leading to enhanced overall clinical results [83]. Integration of genomics data with artificial intelligence and CRISPR-based interventions still paces the advances toward personalized and precision medicine [84].

### **5. Clinical Applications**

#### **5.1 Cancerized Therapy**

CRISPR serves as an enhancement for CAR-T cell therapy to make immune cells effectively target and destroy cancer. It also helps dislodge oncogenes and, thereby, individualized cancer therapy [85].

#### **5.2 Rare Genetic Disorders**

Gene therapy and CRISPR stand for a cure for diseases like sickle cell disease and cystic fibrosis by correcting faulty genes, restoring the normal functioning of proteins, and averting symptoms of the disease [86].

#### **5.3 Neurological Disorders**

Huntington's disease, ALS, and other neurodegenerative disorders that are targeted on such famous genes are being examined through CRISPR path [87].

#### **5.4 Infectious Diseases**

The dissection of CRISPR provides the backbone for the upcoming antiviral agents targeting HIV and various other members of the viral family by directly targeting and disabling their genetic materials to achieve a possible functional cure [88].

### **6. Challenges and Ethical Considerations**

### 6.1 Off-Target Effects and Safety Concerns

Off-target mutations hassle CRISPR-mediated gene editing most because they can lead to unsafe modifications and also recreational safety concerns [89]. Their botheration includes immunological response, carcinomas, or novel genetic diseases [90]. Researchers are thus working up Cas9 variants along with advanced bioinformatics tools, which hold a promise for improvement in specificity while lowering risk [91].

### 6.2. Delivery Efficacy and Immunogenicity

It is the delivery of CRISPR components along with gene therapy vectors to the intended cells, which is again one of the crucial challenges that researchers are facing on in biology [92]. A few benefits in the use of viral vectors (AAV, lentivirus) have already been shown, but some concerns have been raised due to potential uncontrolled immune response that might limit treatment efficiency [93]. By contrast, electroporation and lipid nanoparticles have been found to be more effective but less immunogenic, hence promising as a nonviral delivery method [94]. More complications like continuous gene expression and accurate targeting will be obvious for this approach [95].

### 6.3. Socio-Legal and Ethical Concerns

Eventually, these changes result in genetic disparity and unintentional rupture, in turn causing moral issues arising from genome editing and designer babies [96]. In enlightening this, agencies like that of the FDA and EMA have put strict regulations on gene therapies to ensure safety and ethical consideration in their therapeutic application [97]. All other societal issues are

then public acceptance and fair access by patients to the treatment[98].

## 7. Future Perspectives

### 7.1. Emerging CRISPR Technologies (Base Editing, Epigenome Editing)

CRISPR is advancing past classic gene editing and instituting base editing and epigenome editing that are truly refined and have less collateral damage [99]. Base editing allows for the changing of one DNA letter with no concomitant cutting of the strand: it is amazing for the fixing of point mutations in the widespread sickle cell anemia [100]. Epigenome editing expresses modulation of gene activities without changing the actual sequence to provide possible therapeutic pathways for disorders associated with abnormal gene expression, like cancer and neurological diseases [101].

### 7.2. Smarter and Safer Delivery Methods

The safe and effective delivery of these gene-editing tools into relevant cells is a major challenge[102]. Lipid nanoparticles (LNPs)—the same delivery system used in the mRNA COVID-19 vaccines—have now developed as non-viral vectors that deliver CRISPR [103]. This method elicits less immune response. Although Adeno-associated viruses (AAVs) remain a common option for gene therapy, scientists are modifying them to minimize their risk of eliciting an immune response and improve their capacity to target tissues [104]. Other techniques, such as electroporation and biodegradable carriers, are currently being explored to provide gene delivery with higher precision [105].

### 7.3. The Future of Drug Development with CRISPR



CRISPR and gene therapy are ushering in the era of personal applications and treatments that will last in time [106]. They are capable of targeting not just symptoms but also the genetic problematic of the disease [107]. With AI and machine learning, scientists can further refine which genes to target and further develop their target ideas accordingly [108]. The next shift is in vivo gene editing, in which CRISPR tools are injected directly into a patient's body with the aim of real-time gene correction [109]. Expanding on the impact on health care, CRISPR is beginning to be tested for application beyond rare diseases: with autoimmune diseases, aging disease, and regenerative medicine [110].

## 8. Conclusion

Gene therapy, CRISPR, and other technologies have revolutionized medication development by precisely altering genes to treat genetic abnormalities, cancer, and other ailments. CRISPR allows for focused genome editing, whereas gene therapy adds, changes, or removes genes to fix genetic flaws. Personalized medicine in disease modeling and advancements in drug development were among them.

Some of the FDA-approved gene therapies-Luxturna, Zolgensma, and CAR-T cell therapies-Kymriah, Yescarta-now shine brightly in the landscape of clinical successes. Emerging new variants such as Cas12, Cas13, base editing, and prime editing broaden the CRISPR toolbox and increasingly expedite drug discovery via target identification, drug screening, and precision medicine.

However, there is still an off-target effect, delivery efficacy, immunogenicity, and

ethical challenges. These are some aspects that would warrant intense scrutiny during research. Such future modern medicine can hype an AI, bioinformatics, and genetic tool marriage to safe and effective treatment individualized for each patient.

## 9. References

1. Roemer K, Friedmann T. Concepts and strategies for human gene therapy. *European journal of biochemistry*. 1992 Sep;208(2):211-25. <https://doi.org/10.1111/j.1432-1033.1992.tb17176.x>
2. Martin PA, Turkmendag I. Thinking the unthinkable: how did human germline genome editing become ethically acceptable?. *New Genetics and Society*. 2021 Oct 2;40(4):384-405. <https://doi.org/10.1080/14699915.2021.1932451>
3. Hajebi S, Yousefiasl S, Rahimmanesh I, Dahim A, Ahmadi S, Kadumudi FB, Rahgozar N, Amani S, Kumar A, Kamrani E, Rabiee M. Genetically Engineered Viral Vectors and Organic-Based Non-Viral Nanocarriers for Drug Delivery Applications. *Advanced healthcare materials*. 2022 Oct;11(20):2201583. <https://doi.org/10.1002/adhm.202201583>
4. Boudes PF. Gene therapy as a new treatment option for inherited monogenic diseases. *European journal of internal medicine*. 2014 Jan 1;25(1):31-6. <https://doi.org/10.1016/j.ejim.2013.09.009>
5. Soroudi S, Jaafari MR, Arabi L. Lipid nanoparticle (LNP) mediated mRNA

- delivery in cardiovascular diseases: Advances in genome editing and CAR T cell therapy. *Journal of Controlled Release*. 2024 Aug 1;372:113-40. <https://doi.org/10.1016/j.jconrel.2024.06.023>
6. Lee TL, Sawai T. Navigating equity in global access to genome therapy expanding access to potentially transformative therapies and benefiting those in need requires global policy changes. *Frontiers in Genetics*. 2024 Apr 4;15:1381172. <https://doi.org/10.3389/fgene.2024.1381172>
  7. Sayed N, Allawadhi P, Khurana A, Singh V, Navik U, Pasumarthi SK, Khurana I, Banothu AK, Weiskirchen R, Bharani KK. Gene therapy: Comprehensive overview and therapeutic applications. *Life sciences*. 2022 Apr 1;294:120375. <https://doi.org/10.1016/j.lfs.2022.120375>
  8. Sayed N, Allawadhi P, Khurana A, Singh V, Navik U, Pasumarthi SK, Khurana I, Banothu AK, Weiskirchen R, Bharani KK. Gene therapy: Comprehensive overview and therapeutic applications. *Life sciences*. 2022 Apr 1;294:120375. <https://doi.org/10.1016/j.lfs.2022.120375>
  9. Wachapatthana U, Thanupran T. Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR): A Novel Genomic Modifying. <https://doi.org/10.52403/ijshr.20210744>
  10. Abavisani M, Faraji N, Faraji S, Ebadpour N, Kesharwani P, Sahebkar A. A comprehensive review on utilizing CRISPR/Cas system for microbiome modification. *Biochemical Engineering Journal*. 2024 Jul 26:109443. <https://doi.org/10.1016/j.bej.2024.109443>
  11. Filippova J, Matveeva A, Zhuravlev E, Stepanov G. Guide RNA modification as a way to improve CRISPR/Cas9-based genome-editing systems. *Biochimie*. 2019 Dec 1;167:49-60. <https://doi.org/10.1016/j.biochi.2019.09.003>
  12. Filippova J, Matveeva A, Zhuravlev E, Stepanov G. Guide RNA modification as a way to improve CRISPR/Cas9-based genome-editing systems. *Biochimie*. 2019 Dec 1;167:49-60. <https://doi.org/10.1016/j.biochi.2019.09.003>
  13. Goswami R, Subramanian G, Silayeva L, Newkirk I, Doctor D, Chawla K, Chattopadhyay S, Chandra D, Chilukuri N, Betapudi V. Gene therapy leaves a vicious cycle. *Frontiers in oncology*. 2019 Apr 24;9:297. <https://doi.org/10.3389/fonc.2019.00297>
  14. Bhardwaj A, Nain V. TALENs—an indispensable tool in the era of CRISPR: a mini review. *Journal of Genetic Engineering and Biotechnology*. 2021 Dec 1;19(1):125. <https://doi.org/10.1186/s43141-021-00225-z>

15. Malakar D, Malik HN, Kumar D, Saini S, Sharma V, Fatima S, Bajwa KK, Kumar S. Stem cells: a potential regenerative medicine for treatment of diseases. In *Advances in Animal Genomics 2021 Jan 1* (pp. 33-48). Academic Press. <https://doi.org/10.1016/B978-0-12-820595-2.00003-5>
16. Powell R. In genes we trust: germline engineering, eugenics, and the future of the human genome. In *The Journal of Medicine and Philosophy: A Forum for Bioethics and Philosophy of Medicine 2015 Dec 1* (Vol. 40, No. 6, pp. 669-695). Journal of Medicine and Philosophy Inc.. <https://doi.org/10.1093/jmp/jhv025>
17. Azeez SS, Hamad RS, Hamad BK, Shekha MS, Bergsten P. Advances in CRISPR-Cas technology and its applications: Revolutionising precision medicine. *Frontiers in Genome Editing. 2024 Dec 12*;6:1509924. <https://doi.org/10.3389/fgeed.2024.1509924>
18. Duffy DJ. Problems, challenges and promises: perspectives on precision medicine. *Briefings in bioinformatics. 2016 May 1*;17(3):494-504. <https://doi.org/10.1093/bib/bbv060>
19. Lino CA, Harper JC, Carney JP, Timlin JA. Delivering CRISPR: a review of the challenges and approaches. *Drug delivery. 2018 Jan 1*;25(1):1234-57. <https://doi.org/10.1080/10717544.2018.1474964>
20. Moore JD. The impact of CRISPR-Cas9 on target identification and validation. *Drug discovery today. 2015 Apr 1*;20(4):450-7. <https://doi.org/10.1016/j.drudis.2014.12.016>
21. Zarei A, Razban V, Hosseini SE, Tabei SM. Creating cell and animal models of human disease by genome editing using CRISPR/Cas9. *The journal of gene medicine. 2019 Apr*;21(4):e3082. <https://doi.org/10.1002/jgm.3082>
22. Ratti E, Trist D. Continuing evolution of the drug discovery process in the pharmaceutical industry. *Pure and applied chemistry. 2001 Jan 1*;73(1):67-75. <https://doi.org/10.1351/pac200173010067>
23. Drost J, Van Boxtel R, Blokzijl F, Mizutani T, Sasaki N, Sasselli V, de Ligt J, Behjati S, Grolleman JE, van Wezel T, Nik-Zainal S. Use of CRISPR-modified human stem cell organoids to study the origin of mutational signatures in cancer. *Science. 2017 Oct 13*;358(6360):234-8. <https://doi.org/10.1126/science.aao3130>
24. Zhang B. CRISPR/Cas gene therapy. *Journal of cellular physiology. 2021 Apr*;236(4):2459-81. <https://doi.org/10.1002/jcp.30064>
25. Badwal AK, Singh S. A comprehensive review on the current status of CRISPR based clinical trials for rare diseases. *International Journal of Biological Macromolecules. 2024 Jul 25*;134097. <https://doi.org/10.1016/j.ijbiomac.2024.134097>

26. Zhang B. CRISPR/Cas gene therapy. *Journal of cellular physiology*. 2021 Apr;236(4):2459-81.  
<https://doi.org/10.1002/jcp.30064>
27. Bouard D, Alazard-Dany N, Cosset FL. Viral vectors: from virology to transgene expression. *British journal of pharmacology*. 2009 May;157(2):153-65.  
<https://doi.org/10.1038/bjp.2008.349>
28. Shahryari A, Burtscher I, Nazari Z, Lickert H. Engineering gene therapy: advances and barriers. *Advanced Therapeutics*. 2021 Sep;4(9):2100040.  
<https://doi.org/10.1002/adtp.202100040>
29. Trapani I, Puppo A, Auricchio A. Vector platforms for gene therapy of inherited retinopathies. *Progress in retinal and eye research*. 2014 Nov 1;43:108-28.  
<https://doi.org/10.1016/j.preteyeres.2014.08.001>
30. Sharma D, Arora S, Singh J, Layek B. A review of the tortuous path of nonviral gene delivery and recent progress. *International journal of biological macromolecules*. 2021 Jul 31;183:2055-73.  
<https://doi.org/10.1016/j.ijbiomac.2021.05.192>
31. Kanvinde S, Kulkarni T, Deodhar S, Bhattacharya D, Dasgupta A. Non-viral vectors for delivery of nucleic acid therapies for cancer. *BioTech*. 2022 Mar 7;11(1):6.  
<https://doi.org/10.3390/biotech1101006>
32. Šimčíková M, Prather KL, Prazeres DM, Monteiro GA. Towards effective non-viral gene delivery vector. *Biotechnology and Genetic Engineering Reviews*. 2015 Jul 3;31(1-2):82-107.  
<https://doi.org/10.1080/02648725.2016.1178011>
33. Lammers T, Kiessling F, Hennink WE, Storm G. Drug targeting to tumors: principles, pitfalls and (pre-) clinical progress. *Journal of controlled release*. 2012 Jul 20;161(2):175-87.  
<https://doi.org/10.1016/j.jconrel.2011.09.063>
34. Ginocchio VM, Ferla R, Auricchio A, Brunetti-Pierri N. Current status on clinical development of adeno-associated virus-mediated liver-directed gene therapy for inborn errors of metabolism. *Human Gene Therapy*. 2019 Oct 1;30(10):1204-10.  
<https://doi.org/10.1089/hum.2019.151>
35. Dhurandhar D, Sahoo NK, Mariappan I, Narayanan R. Gene therapy in retinal diseases: A review. *Indian Journal of Ophthalmology*. 2021 Sep 1;69(9):2257-65.  
10.4103/ijo.IJO\_3117\_20
36. Dhurandhar D, Sahoo NK, Mariappan I, Narayanan R. Gene therapy in retinal diseases: A review. *Indian Journal of Ophthalmology*. 2021 Sep 1;69(9):2257-65.  
10.4103/ijo.IJO\_3117\_20
37. Ponomarev AS, Chulpanova DS, Yanygina LM, Solovyeva VV, Rizvanov AA. Emerging gene therapy approaches in the management of spinal muscular atrophy (SMA): An overview of clinical trials and patent

- landscape. International Journal of Molecular Sciences. 2023 Sep 6;24(18):13743.  
<https://doi.org/10.3390/ijms241813743>
38. Aslesh T, Yokota T. Restoring SMN expression: an overview of the therapeutic developments for the treatment of spinal muscular atrophy. Cells. 2022 Jan 26;11(3):417.  
<https://doi.org/10.3390/cells11030417>
39. Ali S, Kjekken R, Niederlaender C, Markey G, Saunders TS, Opsata M, Moltu K, Bremnes B, Grønevik E, Muusse M, Håkonsen GD. The European medicines agency review of Kymriah (Tisagenlecleucel) for the treatment of acute lymphoblastic leukemia and diffuse large B-cell lymphoma. The oncologist. 2020 Feb 1;25(2):e321-7.  
<https://doi.org/10.1634/theoncologist.2019-0233>
40. Schmitt TM, Stromnes IM, Chapuis AG, Greenberg PD. New strategies in engineering T-cell receptor gene-modified T cells to more effectively target malignancies. Clinical cancer research. 2015 Dec 1;21(23):5191-7.  
<https://doi.org/10.1158/1078-0432.CCR-15-0860>
41. Mohanty R, Chowdhury CR, Arega S, Sen P, Ganguly P, Ganguly N. CAR T cell therapy: A new era for cancer treatment. Oncology reports. 2019 Dec 1;42(6):2183-95.  
<https://doi.org/10.3892/or.2019.7335>
42. Mohanty R, Chowdhury CR, Arega S, Sen P, Ganguly P, Ganguly N. CAR T cell therapy: A new era for cancer treatment. Oncology reports. 2019 Dec 1;42(6):2183-95.  
<https://doi.org/10.3892/or.2019.7335>
43. Khvorost D, Kendall B, Jazirehi AR. Immunotherapy of hematological malignancies of human B-cell origin with CD19 CAR T lymphocytes. Cells. 2024 Apr 9;13(8):662.  
<https://doi.org/10.3390/cells13080662>
44. Bogacz A, Bukowska A, Bukowska M, Olbromski K, Łaba A, Klupieć R, Jopek K. Modern immunotherapy using CAR-T cells in haemat-oncology and solid tumors. Acta Haematologica Polonica. 2024;55(1):34-41.  
<https://doi.org/10.5603/ahp.97189>
45. Szlasa W, Dybko J. Current status of bispecific antibodies and CAR-T therapies in multiple myeloma. International Immunopharmacology. 2024 Jun 15;134:112043.  
<https://doi.org/10.1016/j.intimp.2024.112043>
46. Alomari M, Kunacheewa C, Manasanch EE. The role of soluble B cell maturation antigen as a biomarker in multiple myeloma. Leukemia & Lymphoma. 2023 Jan 28;64(2):261-72.  
<https://doi.org/10.1080/10428194.2022.2133540>
47. Yıldırım S. Gene Therapy Products Approved in 2022. Gene Editing. 2022 Dec 25;3(3):9-13.  
10.29228/genediting.67056



48. Giommetti A, Papanikolaou E. Advancements in Hematopoietic Stem Cell Gene Therapy: A Journey of Progress for Viral Transduction. *Cells*. 2024 Jun 15;13(12):1039. <https://doi.org/10.3390/cells13121039>
49. Soroka AB, Feoktistova SG, Mityaeva ON, Volchkov PY. Gene therapy approaches for the treatment of hemophilia B. *International Journal of Molecular Sciences*. 2023 Jun 28;24(13):10766. <https://doi.org/10.3390/ijms241310766>
50. Anguela XM, High KA. Hemophilia B and gene therapy: A new chapter with etranacogene dezaparvovec. *Blood Advances*. 2024 Apr 9;8(7):1796-803. <https://doi.org/10.1182/bloodadvances.2023010511>
51. Bueren JA, Auricchio A. Advances and challenges in the development of gene therapy medicinal products for rare diseases. *Human Gene Therapy*. 2023 Sep 1;34(17-18):763-75. <https://doi.org/10.1089/hum.2023.152>
52. Bueren JA, Auricchio A. Advances and challenges in the development of gene therapy medicinal products for rare diseases. *Human Gene Therapy*. 2023 Sep 1;34(17-18):763-75. <https://doi.org/10.1089/hum.2023.152>
53. Yan S, McDade C, Thiruvillakkat K, Rouse R, Sivamurthy K, Wilson M. Analysis of long-term clinical and cost impact of etranacogene dezaparvovec for the treatment of hemophilia B population in the United States. *Journal of Medical Economics*. 2024 Dec 31;27(1):758-65. <https://doi.org/10.1080/13696998.2024.2351762>
54. Janik E, Niemcewicz M, Ceremuga M, Krzowski L, Saluk-Bijak J, Bijak M. Various aspects of a gene editing system—crispr-cas9. *International journal of molecular sciences*. 2020 Dec 16;21(24):9604. <https://doi.org/10.3390/ijms21249604>
55. Doudna JA, Charpentier E. The new frontier of genome engineering with CRISPR-Cas9. *Science*. 2014 Nov 28;346(6213):1258096. <https://doi.org/10.1126/science.1258096>
56. Burma S, Chen BP, Chen DJ. Role of non-homologous end joining (NHEJ) in maintaining genomic integrity. *DNA repair*. 2006 Sep 8;5(9-10):1042-8. <https://doi.org/10.1016/j.dnarep.2006.05.026>
57. Karimian A, Azizian K, Parsian H, Rafieian S, Shafiei-Irannejad V, Kheyrollah M, Yousefi M, Majidinia M, Yousefi B. CRISPR/Cas9 technology as a potent molecular tool for gene therapy. *Journal of cellular physiology*. 2019 Aug;234(8):12267-77. <https://doi.org/10.1002/jcp.27972>
58. Manghwar H, Li B, Ding X, Hussain A, Lindsey K, Zhang X, Jin S. CRISPR/Cas systems in genome editing: methodologies and tools for sgRNA design, off-target evaluation, and strategies to mitigate off-target effects. *Advanced science*. 2020

- Mar;7(6):1902312.  
10.1002/advs.201902312
59. Son H. Harnessing CRISPR/Cas Systems for DNA and RNA Detection: Principles, Techniques, and Challenges. *Biosensors*. 2024 Sep 26;14(10):460.  
<https://doi.org/10.3390/bios14100460>
  60. Kordyś M, Sen R, Warkocki Z. Applications of the versatile CRISPR-Cas13 RNA targeting system. *Wiley Interdisciplinary Reviews: RNA*. 2022 May;13(3):e1694.  
<https://doi.org/10.1002/wrna.1694>
  61. Kantor A, McClements ME, MacLaren RE. CRISPR-Cas9 DNA base-editing and prime-editing. *International journal of molecular sciences*. 2020 Aug 28;21(17):6240.  
<https://doi.org/10.3390/ijms21176240>
  62. Petrova IO, Smirnikhina SA. The development, optimization and future of prime editing. *International Journal of Molecular Sciences*. 2023 Dec 1;24(23):17045.  
<https://doi.org/10.3390/ijms242317045>
  63. Moore JD. The impact of CRISPR-Cas9 on target identification and validation. *Drug discovery today*. 2015 Apr 1;20(4):450-7.  
<https://doi.org/10.1016/j.drudis.2014.12.016>
  64. Thomford NE, Senthane DA, Rowe A, Munro D, Seele P, Maroyi A, Dzobo K. Natural products for drug discovery in the 21st century: innovations for novel drug discovery. *International journal of molecular sciences*. 2018 May 25;19(6):1578.  
<https://doi.org/10.3390/ijms19061578>
  65. Nagini S. Breast cancer: current molecular therapeutic targets and new players. *Anti-Cancer Agents in Medicinal Chemistry-Anti-Cancer Agents*. 2017 Feb 1;17(2):152-63.  
<https://doi.org/10.2174/1871520616666160502122724>
  66. Rodríguez-Frías F, Quer J, Tabernero D, Cortese MF, Garcia-Garcia S, Rando-Segura A, Pumarola T. Microorganisms as shapers of human civilization, from pandemics to even our genomes: villains or friends? A Historical approach. *Microorganisms*. 2021 Dec 6;9(12):2518.  
<https://doi.org/10.3390/microorganisms9122518>
  67. Szymański P, Markowicz M, Mikiciuk-Olasik E. Adaptation of high-throughput screening in drug discovery – toxicological screening tests. *International journal of molecular sciences*. 2011 Dec 29;13(1):427-52.  
<https://doi.org/10.3390/ijms13010427>
  68. Naithani U, Guleria V. Integrative computational approaches for discovery and evaluation of lead compound for drug design. *Frontiers in Drug Discovery*. 2024 Apr 5;4:1362456.  
<https://doi.org/10.3389/fddsv.2024.1362456>
  69. Marques L, Costa B, Pereira M, Silva A, Santos J, Saldanha L, Silva I,

- Magalhães P, Schmidt S, Vale N. Advancing precision medicine: a review of innovative in silico approaches for drug development, clinical pharmacology and personalized healthcare. *Pharmaceutics*. 2024 Feb 27;16(3):332. <https://doi.org/10.3390/pharmaceutics16030332>
70. Haq EU, Yousaf A, Sardar M, Irshad A, Basharat Z, Ali A, Gillani SM, Arsalan M. Uncovering Genetic Interactions: CRISPR-Mediated Gene Knockouts and Activations in Understanding Complex Diseases. *Pak-Euro Journal of Medical and Life Sciences*. 2024 Dec 15;7(Special 2):S211-20. <https://doi.org/10.31580/pjmls.v7isp2.3164>
  71. Lu Q, Livi GP, Modha S, Yusa K, Macarrón R, Dow DJ. Applications of CRISPR genome editing technology in drug target identification and validation. *Expert opinion on drug discovery*. 2017 Jun 3;12(6):541-52. <https://doi.org/10.1080/17460441.2017.1317244>
  72. Venkatesh S, Lipper RA. Role of the development scientist in compound lead selection and optimization. *Journal of pharmaceutical sciences*. 2000 Feb 1;89(2):145-54. [https://doi.org/10.1002/\(SICI\)1520-6017\(200002\)89:2%3C145::AID-IPS2%3E3.0.CO;2-6](https://doi.org/10.1002/(SICI)1520-6017(200002)89:2%3C145::AID-IPS2%3E3.0.CO;2-6)
  73. Sajjad H, Imtiaz S, Noor T, Siddiqui YH, Sajjad A, Zia M. Cancer models in preclinical research: A chronicle review of advancement in effective cancer research. *Animal models and experimental medicine*. 2021 Jun;4(2):87-103. <https://doi.org/10.1002/ame2.12165>
  74. Macarrón Palacios A, Korus P, Wilkens BG, Heshmatpour N, Patnaik SR. Revolutionizing in vivo therapy with CRISPR/Cas genome editing: breakthroughs, opportunities and challenges. *Frontiers in Genome Editing*. 2024 Feb 1;6:1342193. <https://doi.org/10.3389/fgeed.2024.1342193>
  75. Behl T, Kaur I, Sehgal A, Singh S, Albarrati A, Albratty M, Najmi A, Meraya AM, Bungau S. The road to precision medicine: Eliminating the “One Size Fits All” approach in Alzheimer’s disease. *Biomedicine & Pharmacotherapy*. 2022 Sep 1;153:113337. <https://doi.org/10.1016/j.biopha.2022.113337>
  76. Chanchal DK, Chaudhary JS, Kumar P, Agnihotri N, Porwal P. CRISPR-based therapies: revolutionizing drug development and precision medicine. *Current Gene Therapy*. 2024 Jun 1;24(3):193-207. <https://doi.org/10.2174/0115665232275754231204072320>
  77. Zhang B. CRISPR/Cas gene therapy. *Journal of cellular physiology*. 2021 Apr;236(4):2459-81. <https://doi.org/10.1002/jcp.30064>
  78. Gardner RV. Sickle cell disease: advances in treatment. *Ochsner journal*. 2018 Dec 21;18(4):377-89. <https://doi.org/10.31486/toj.18.0076>

79. Ye J, Li D, Jie Y, Luo H, Zhang W, Qiu C. Exosome-based nanoparticles and cancer immunotherapy. *Biomedicine & Pharmacotherapy*. 2024 Oct 1;179:117296. <https://doi.org/10.1016/j.biopha.2024.117296>
80. Dunbar CE, High KA, Joung JK, Kohn DB, Ozawa K, Sadelain M. Gene therapy comes of age. *Science*. 2018 Jan 12;359(6372):eaan4672. <https://doi.org/10.1126/science.aan4672>
81. Goetz LH, Schork NJ. Personalized medicine: motivation, challenges, and progress. *Fertility and sterility*. 2018 Jun 1;109(6):952-63. <https://doi.org/10.1016/j.fertnstert.2018.05.006>
82. Jensen TL, Gøtzsche CR, Woldbye DP. Current and future prospects for gene therapy for rare genetic diseases affecting the brain and spinal cord. *Frontiers in molecular neuroscience*. 2021 Oct 6;14:695937. <https://doi.org/10.3389/fnmol.2021.695937>
83. Macarrón Palacios A, Korus P, Wilkens BG, Heshmatpour N, Patnaik SR. Revolutionizing in vivo therapy with CRISPR/Cas genome editing: breakthroughs, opportunities and challenges. *Frontiers in Genome Editing*. 2024 Feb 1;6:1342193. <https://doi.org/10.3389/fgeed.2024.1342193>
84. Azeez SS, Hamad RS, Hamad BK, Shekha MS, Bergsten P. Advances in CRISPR-Cas technology and its applications: Revolutionising precision medicine. *Frontiers in Genome Editing*. 2024 Dec 12;6:1509924. <https://doi.org/10.3389/fgeed.2024.1509924>
85. Amiri M, Moaveni AK, Majidi Zolbin M, Shademan B, Nourazarian A. Optimizing cancer treatment: the synergistic potential of CAR-T cell therapy and CRISPR/Cas9. *Frontiers in Immunology*. 2024 Nov 8;15:1462697. <https://doi.org/10.3389/fimmu.2024.1462697>
86. Kotagama OW, Jayasinghe CD, Abeysinghe T. Era of genomic medicine: a narrative review on CRISPR technology as a potential therapeutic tool for human diseases. *BioMed research international*. 2019;2019(1):1369682. <https://doi.org/10.1155/2019/1369682>
87. Akyuz E, Aslan FS, Gokce E, Ilmaz O, Topcu F, Kakac S. Extracellular vesicle and CRISPR gene therapy: Current applications in Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis, and Huntington's disease. *European Journal of Neuroscience*. 2024 Oct;60(8):6057-90. <https://doi.org/10.1111/ejn.16541>
88. Banda A, Impomeni O, Singh A, Baloch AR, Hu W, Jaijyan DK. Precision in Action: The Role of Clustered Regularly Interspaced Short Palindromic Repeats/Cas in Gene Therapies. *Vaccines*. 2024 Jun 7;12(6):636.

- <https://doi.org/10.3390/vaccines12060636>
89. Piergentili R, Del Rio A, Signore F, Umani Ronchi F, Marinelli E, Zaami S. CRISPR-Cas and its wide-ranging applications: From human genome editing to environmental implications, technical limitations, hazards and bioethical issues. *Cells*. 2021 Apr 21;10(5):969. <https://doi.org/10.3390/cells10050969>
  90. MacDougall M, El-Hajj Sleiman J, Beauchemin P, Rangachari M. SARS-CoV-2 and multiple sclerosis: potential for disease exacerbation. *Frontiers in Immunology*. 2022 Apr 22;13:871276. <https://doi.org/10.3389/fimmu.2022.871276>
  91. Dixit S, Kumar A, Srinivasan K, Vincent PD, Ramu Krishnan N. Advancing genome editing with artificial intelligence: opportunities, challenges, and future directions. *Frontiers in Bioengineering and Biotechnology*. 2024 Jan 8;11:1335901. <https://doi.org/10.3389/fbioe.2023.1335901>
  92. Uddin F, Rudin CM, Sen T. CRISPR gene therapy: applications, limitations, and implications for the future. *Frontiers in oncology*. 2020 Aug 7;10:1387. <https://doi.org/10.3389/fonc.2020.01387>
  93. Tomanin R, Scarpa M. Why do we need new gene therapy viral vectors? Characteristics, limitations and future perspectives of viral vector transduction. *Current gene therapy*. 2004 Dec 1;4(4):357-72. <https://doi.org/10.2174/1566523043346011>
  94. Harris E, Elmer JJ. Optimization of electroporation and other non-viral gene delivery strategies for T cells. *Biotechnology progress*. 2021 Jan;37(1):e3066. <https://doi.org/10.1002/btpr.3066>
  95. Anomaly J. Creating future people: The ethics of genetic enhancement. Routledge; 2020 Jan 28. <https://doi.org/10.4324/9781003014805>
  96. Kalidasan V, Theva Das K. Advancing Precision Medicine with Gene and Cell Therapy in Malaysia: Ethical, Legal, and Social Implications. *Human Gene Therapy*. 2024 Jan 1;35(1-2):9-25. <https://doi.org/10.1089/hum.2023.139>
  97. Kalidasan V, Theva Das K. Advancing Precision Medicine with Gene and Cell Therapy in Malaysia: Ethical, Legal, and Social Implications. *Human Gene Therapy*. 2024 Jan 1;35(1-2):9-25. <https://doi.org/10.1089/hum.2023.139>
  98. 139 Lawler C, Gu L, Howard LE, Branche B, Wiggins E, Srinivasan A, Foster ML, Klaassen Z, De Hoedt AM, Gingrich JR, Theodorescu D. The impact of the social construct of race on outcomes among bacille Calmette- Guérin- treated patients with high- risk non- muscle-invasive bladder cancer in an equal- access setting. *Cancer*. 2021 Nov 1;127(21):3998-4005. <https://doi.org/10.1002/cncr.33792>



99. Chira S, Nutu A, Isacescu E, Bica C, Pop L, Ciocan C, Berindan-Neagoe I. Genome editing approaches with CRISPR/Cas9 for cancer treatment: Critical appraisal of preclinical and clinical utility, challenges, and future research. *Cells*. 2022 Sep 6;11(18):2781. <https://doi.org/10.3390/cells11182781>
100. Nidhi S, Anand U, Oleksak P, Tripathi P, Lal JA, Thomas G, Kuca K, Tripathi V. Novel CRISPR-Cas systems: an updated review of the current achievements, applications, and future research perspectives. *International journal of molecular sciences*. 2021 Mar 24;22(7):3327. <https://doi.org/10.3390/ijms22073327>
101. Duarte F, Déglon N. Genome editing for CNS disorders. *Frontiers in neuroscience*. 2020 Oct 22;14:579062. <https://doi.org/10.3389/fnins.2020.579062>
102. Taha EA, Lee J, Hotta A. Delivery of CRISPR-Cas tools for in vivo genome editing therapy: Trends and challenges. *Journal of Controlled Release*. 2022 Feb 1;342:345-61. <https://doi.org/10.1016/j.jconrel.2022.01.013>
103. Yang L, Gong L, Wang P, Zhao X, Zhao F, Zhang Z, Li Y, Huang W. Recent advances in lipid nanoparticles for delivery of mRNA. *Pharmaceutics*. 2022 Dec 1;14(12):2682. <https://doi.org/10.3390/pharmaceutics14122682>
104. Nidetz NF, McGee MC, Longping VT, Li C, Cong L, Li Y, Huang W. Adeno-associated viral vector-mediated immune responses: Understanding barriers to gene delivery. *Pharmacology & therapeutics*. 2020 Mar 1;207:107453. <https://doi.org/10.1016/j.pharmthera.2019.107453>
105. Shi J, Ma Y, Zhu J, Chen Y, Sun Y, Yao Y, Yang Z, Xie J. A review on electroporation-based intracellular delivery. *Molecules*. 2018 Nov 21;23(11):3044. <https://doi.org/10.3390/molecules23113044>
106. Goswami R, Subramanian G, Silayeva L, Newkirk I, Doctor D, Chawla K, Chattopadhyay S, Chandra D, Chilukuri N, Betapudi V. Gene therapy leaves a vicious cycle. *Frontiers in oncology*. 2019 Apr 24;9:297. <https://doi.org/10.3389/fonc.2019.00297>
107. Watts JK, Corey DR. Silencing disease genes in the laboratory and the clinic. *The Journal of pathology*. 2012 Jan;226(2):365-79. <https://doi.org/10.1002/path.2993>
108. Sarkar C, Das B, Rawat VS, Wahlang JB, Nongpiur A, Tiewsoh I, Lyngdoh NM, Das D, Bidarolli M, Sony HT. Artificial intelligence and machine learning technology driven modern drug discovery and development. *International Journal of Molecular Sciences*. 2023 Jan 19;24(3):2026. <https://doi.org/10.3390/ijms24032026>
109. Lohia A, Sahel DK, Salman M, Singh V, Mariappan I, Mittal A, Chitkara D.

Delivery strategies for CRISPR/Cas genome editing tool for retinal dystrophies: challenges and opportunities. Asian Journal of Pharmaceutical Sciences. 2022 Mar 1;17(2):153-76.

<https://doi.org/10.1016/j.ajps.2022.02.001>

110. Greco F, Cosentino M, Marino F. The Italian breakthrough in CRISPR trials for rare diseases: a focus on beta-thalassemia and sickle cell disease treatment. Frontiers in Medicine. 2024 Feb 15;11:1356578.

<https://doi.org/10.3389/fmed.2024.1356578>

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